

Instructions for Use

MYCOMOUNT™

Cat. no. MM40	MycoMount™	40 mounting strips/pkg
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INTENDED USE

Hardy Diagnostics MycoMount™ is a device used to facilitate the preparation of fungal wet mounts for microscopic examination. It allows easy access to tubes and bottles and eliminates the need for "tease mounts" and "scotch tape preps."

SUMMARY

MycoMount™ is a clear, narrow strip of flexible material with a pressure sensitive adhesive at the tip. It is used to pick up fruiting structures located on the top of fungal colonies. The long flexible handle allows easy access to tubes and bottles and keeps potentially infectious spores at a distance, which can be a problem with traditional "scotch tape" preps. A second adhesive pad firmly attaches the strip to the microscope slide. A score mark is located just below the second adhesive strip so that the handle can be easily broken off and discarded. The clarity of the strip allows the fungal elements to be viewed microscopically through the strip in a wet mount.

STORAGE

Storage: Upon receipt store at 15-30°C.

PRECAUTIONS

This product may contain components of animal origin. Certified knowledge of the origin and/or sanitary state of the animals does not guarantee the absence of transmissible pathogenic agents. Therefore, it is recommended that these products be treated as potentially infectious, and handle observing the usual Universal Precautions for blood. Do not ingest, inhale, or allow to come into contact with skin.

This product is for *in vitro* diagnostic use only. It is to be used only by adequately trained and qualified laboratory personnel. Observe approved biohazard precautions and aseptic techniques. All laboratory specimens should be considered infectious and handled according to "standard precautions." Refer to the document "[Guidelines for Isolation Precautions](#)" from the Centers for Disease Control and Prevention.

For additional information regarding specific precautions for the prevention of the transmission of all infectious agents from laboratory instruments and materials, and for recommendations for the management of exposure to infectious disease, refer to CLSI document M29: *Protection of Laboratory Workers from Occupationally Acquired Infections*.

Sterilize all biohazard waste before disposal.

Refer to the document "[Precautions When Using Media](#)" for more information.

PROCEDURE

1. Place a single drop of Lactophenol Cotton Blue Stain (Cat. no. Z68) at the end of a clean glass slide about 3/4 inch from the right end of the slide.
2. Remove one strip from the container. Using forceps, remove the paper from the adhesive on the far end of the strip. (Do not remove the paper in the middle of the strip yet.)
3. Insert the strip into the tube or bottle and press the exposed adhesive tip gently to the culture to pick up the fruiting structures of the mature mold colony.
4. Using forceps, now remove the paper from the middle of the strip.
5. Place the strip on the slide with the culture specimen face down on the lactophenol cotton blue drop. With forceps, press down on the middle adhesive strip to attach the strip to the slide. If desired, the excess portion of the strip can be broken off by bending the strip down at the score mark which is located just below the middle adhesive strip.

INTERPRETATION OF RESULTS

View the wet mount under the microscope at 100X or 400X. Consult listed references for information on microscopic morphology and identification of fungi.⁽¹⁻⁶⁾

LIMITATIONS

Wet mounts are useful in the recognition and presumptive identification of fungi. Additional characteristics including colony morphology and biochemical tests should be used where appropriate for final identification. For further information, consult appropriate references.⁽¹⁻⁶⁾

Failure to find conidia or spores may be due to immature colonies that are too young to produce fruiting structures. Absence of conidia or spores may also be due to the inability of the organism to sporulate on the culture medium used.

Antibiotics in the medium used for cultivation may inhibit or retard sporulation of molds.

MATERIALS REQUIRED BUT NOT PROVIDED

Standard microbiological supplies and equipment such as forceps, slides, stains and microscopes, etc., are not provided.

REFERENCES

1. Isenberg, H.D. *Clinical Microbiology Procedures Handbook*, Vol. I, II & III. American Society for Microbiology, Washington, D.C.
2. Jorgensen., et al. *Manual of Clinical Microbiology*, American Society for Microbiology, Washington, D.C.
3. Kwon-Chung, K.J. and J.E. Bennett. 1992. *Medical Mycology*. Lea and Febiger, Malvern, PA.
4. Larone, D.H. *Medically Important Fungi: A Guide to Identification*, American Society for Microbiology, Washington, D.C.
5. *Cumitech 11: Practical Methods for Culture and Identification of Fungi in the Clinical Microbiology Laboratory*. 1980. American Society for Microbiology, Washington, D.C.
6. St. Germain, Guy, et al. 1996. *Identifying Filamentous Fungi*. Star Publishing Company, Belmont, CA.



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